



## **FINAL REPORT**

**Skin Irritation Test in New Zealand White Rabbits of Taglus PU FLEX Thermoforming Foils as per ISO 10993-23:2021(E).**

**STUDY CONTRACT PARTNER:**

UL India Private Limited

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**UL Project Number: 4790186870**

**TEST FACILITY:**

GLR Laboratories Private Limited,

444, Gokulam Street, Mathur, Chennai - 600 068, Tamil Nadu, India.

**Study No.: 073/437**

**STUDY SPONSOR AND APPLICANT:**

Vedia Solutions

Division of Laxmi Dental Export Pvt Ltd,

103, Akruiti Arcade, J P Road,

Opp A H Wadia School, Andheri West

Mumbai 400053

**REPORT ISSUED DATE: 30 December 2021**



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**FLEX Thermoforming Foils as per ISO 10993-23:2021(E).**

**Study No:**  
**073/437**

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### **PRODUCT NAME:**

**Taglus PU FLEX Thermoforming Foils**

### **STUDY TITLE**

**Skin Irritation Test in New Zealand White Rabbits of**  
**Taglus PU FLEX Thermoforming Foils as per**  
**ISO 10993-23:2021(E)**

**STUDY NUMBER:**  
**073/437**

**TEST FACILITY:**  
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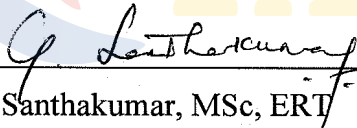
**STUDY DIRECTOR AUTHENTICATION STATEMENT**


**Study No : 073/437**

**Study Title : Skin Irritation Test in New Zealand White Rabbits of Taglus PU**  
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This study was performed in accordance with the mutually agreed study plan and GLR Laboratories Private Limited's standard operating procedures, unless otherwise stated, and the study objective was achieved. I accept overall responsibility for the technical conduct of the study, as well as for the interpretation, analysis, documentation and reporting of results. This report provides a true and accurate record of the results obtained.

This study was performed in compliance with OECD Principles of Good Laboratory Practice\* ENV/MC/CHEM (98)17 (Revised 1997, issued January 1998) and applicable regulatory requirements including the US Food and Drug Administration's GLP regulations, 21 CFR 58 (subparts B to G and J).

  
\_\_\_\_\_  
Mr. G. Santhakumar, MSc, ERT  
Study Director  
GLR Laboratories Private Limited

  
\_\_\_\_\_  
30 Dec 2021  
Study Completion Date

\*The identity and composition of the test item are the responsibilities of the Sponsor.



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**QUALITY ASSURANCE STATEMENT**

**Study No : 073/437**

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The Quality Assurance (QA) of GLR Laboratories Private Limited verified the Study Plan including any amendments, inspected the critical study phases, and audited the raw data and report of this Study as per in-house Standard Operating Procedures (SOPs) for compliance with the OECD Principles of Good Laboratory Practice (as revised in 1997) [ENV/MC/CHEM (98)17], and for compliance with relevant regulatory requirements.

During the Study, the following study-related inspections/audits were performed on the following dates and reported to the Study Director and Test Facility Management. Besides the below, process and facility inspections were also carried out periodically at this Test Facility by auditor(s) of the QA, as per in-house SOPs, which may have relevance to this study.

<b>S. No.</b>	<b>Type of Inspection</b>	<b>Date of Inspection</b>	<b>Phase(s) of Study Inspected</b>	<b>Date of Reporting to Management, Study Director (Inspection No.)</b>
1	Study Plan Verification	20 November 2021	Draft Study Plan	20 November 2021 (SBI/073/437/001)
2	Study Plan Verification	26 November 2021	Definitive Study Plan	26 November 2021 (SBI/073/437/002)
3	In Life Phase Inspection	06 December 2021	Test Item Extract Application	06 December 2021 (SBI/073/437/003)
4	In Life Phase Inspection	09 December 2021	Grading of Skin Reactions	09 December 2021 (SBI/073/437/004)
5	Report Audit	24 December 2021	Draft Report	24 December 2021 (SBI/073/437/005)
6	Report Audit	30 December 2021	Final Report	30 December 2021 (SBI/073/437/006)



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The QA has determined that the methods, procedures, observations, and reported results are accurately and completely described and that the reported results are based on the Study Plan and the pertinent raw data generated during the course of the Study. The Study Director's GLP compliance statement is supported.

*N. Pull*

*30 DEC 2021*

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Dr. Parthiban Natarajan, PhD, ERT  
Head - Quality Assurance  
Asst. Director, GLR Laboratories Private Limited

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Date





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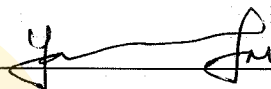
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**TEST FACILITY MANAGEMENT STATEMENT**

**Study No : 073/437**

**Study Title : Skin Irritation Test in New Zealand White Rabbits of Taglus PU**  
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This is to certify that, the Test Facility Management appointed the Study Director and provided all necessary facilities and resources for the proper conduct of this study, in compliance with the Principles of OECD Good Laboratory Practice (GLP), as per the recommendations of the OECD (Council Act [C (97) 186 (Final)]) and as adopted in the procedures promulgated by the National GLP Compliance Monitoring Authority, Government of India.

  
\_\_\_\_\_  
Ms. M. Yaminy, B. Com, (MBA)  
Deputy Test Facility Management  
GLR Laboratories Private Limited

30 Dec 2021  
\_\_\_\_\_  
Date



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### **SUMMARY**

Skin irritation potential of Taglus PU FLEX Thermoforming Foils, supplied by Vedia Solutions, was evaluated in male New Zealand White rabbits.

The test item, Taglus PU FLEX Thermoforming Foils is a transparent disk with a diameter of 12.5 cm and thickness less than 0.5 mm. It is a surface device which comes in contact with mucosal membrane. The duration of contact is less than 24 hours (limited).

The whole test item was extracted at a ratio of 6 cm<sup>2</sup>/mL (since the thickness of the test item is less than 0.5 mm) in polar solvent (physiological saline) as well as non-polar solvent (cottonseed oil) at 37 ± 1 °C for 72 h and 02 min. The total surface area of the test item is 111 cm<sup>2</sup> (as calculated in our laboratory). Polar extract was prepared by extracting 111 cm<sup>2</sup> of test item in 18.5 mL of physiological saline. Similarly, non-polar extract was freshly prepared by extracting 111 cm<sup>2</sup> of test item in 18.5 mL of cottonseed oil. Solvent controls were also subjected to the similar extraction conditions. This fulfils the requirement of ISO 10993-12:2012(E) and ISO 10993-12:2021(E).

At the end of extraction, extracts and solvent controls were clear without any colour change or particulates. No changes were observed in the retrieved test item. No additional processing such as filtration, centrifugation, pH adjustments or any other processing were made. The extracts and solvent controls were transferred to sterile containers and stored at room temperature. All extracts and solvent controls were used within 1 h 23 min of preparation and were considered stable during this time.

About 16 h and 55 min prior to application, fur on all the rabbits were closely clipped off their backs, allowing sufficient distance on both sides of the spine for application of test item extracts and solvent controls. The test item extracts (0.5 mL) was loaded on an absorbent gauze patch measuring 6.25 cm<sup>2</sup> (2.5 cm x 2.5 cm) and placed topically on the fur clipped area of six male rabbits (3 animals each for polar and non-polar extracts). The patches were loosely held in contact with the skin by semi-occlusive dressing with means of non-irritant adhesive tape for 4 h. After 4 h the patch was removed. No test item extract residues were observed. The test sites were marked with a non- irritant permanent ink.

Animals were observed for three consecutive days for morbidity, mortality, skin reactions and abnormal clinical signs and symptoms following the patch removal. The skin reactions were visually scored according to ISO 10993-23:2021(E) at 1, 24, 48 and 72 h.

Positive control trials for irritation are carried out periodically at GLR Laboratories Private Limited in New Zealand White rabbits using sodium lauryl sulphate in compliance with regulatory guidelines. The trial completed on 20 October 2021 gave a “moderate irritant”





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reaction. No response was observed in solvent control treated animals. Therefore, the assay was considered valid.

No mortality or morbidity was observed in the experimental animals. A gradual increase in body weight was observed in all the animals at the end of the experiment. No signs of clinical toxicity or overt toxicity was observed in any of the animals. Hence, gross pathology and histopathology were not performed. No local skin irritation was observed at the test site in any of the animals and the primary irritation index was '0'.

Based upon the results obtained in this study and in line with ISO 10993-23:2021(E), the given test item, Taglus PU FLEX Thermoforming Foils, supplied by Vedia Solutions, is considered as a non-irritant under the conditions of the present study.





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## **INTRODUCTION**

Biocompatibility testing is a regulatory requirement for demonstrating the preclinical safety of medical devices. This is evaluated in line with the standard guideline, ISO 10993-1:2018(E), Biological Evaluation of Medical Devices - Part 1, Evaluation and Testing within a Risk Management Process. This standard describes the necessity to select a suitable test method for biocompatibility evaluation.

Skin irritation is a key toxicity endpoint to assess biocompatibility of medical devices. An assessment is made of the potential of the material under test to produce dermal irritation in rabbits following topical application.

## **OBJECTIVE**

To determine the skin irritation potential of the test item in New Zealand white rabbits.

## **STUDY DATES**

Study Start Date	26 November 2021
Experiment Start Date	30 November 2021
Experiment Completion Date	09 December 2021

The study completion date is the date the final report is signed by the Study Director.

## **TEST ITEM DETAILS**

The test item, Taglus PU FLEX Thermoforming Foils was received at GLR Laboratories Private Limited on 13 November 2021 and stored at room temperature (20.1 to 24.1 °C) until used.

The following test item information provided by the sponsor were considered adequate.

Test Item	Taglus PU FLEX Thermoforming Foils
Batch \ Lot No.	12029092-1
Manufacture Date	29 September 2021
Expiry Date	20 September 2024
Appearance	Transparent disk
Ingredients	PU (PolyUrethane)
Temperature Stability	37 °C
Sterility	Non-Sterile



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**CONTROL ITEM DETAILS**

Positive Control	Sodium Lauryl Sulphate (SLS)
Manufacturer	Sigma Aldrich
Batch No.	0000009635
Expiry date	August 2022

Positive control trials for irritation are carried out periodically at GLR Laboratories Private Limited in New Zealand White rabbits using sodium lauryl sulphate in compliance with regulatory guidelines (ISO 10993-10:2010(E), ISO 10993-23:2021(E) and OECD 404:2015). The trial completed on 20 October 2021 gave a “moderate irritant” reaction (Appendix 1).

Solvent Controls	<u>Physiological saline</u>
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Manufacturer	Eurolife Healthcare Pvt. Ltd.
Batch No.	10210671B

Expiry Date	September 2024
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Appearance	Colourless clear liquid
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<u>Cottonseed oil</u>	
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Manufacturer	Sigma-Aldrich
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Lot No.	MKCM9272
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Expiry Date	March 2026
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Appearance	Yellow coloured viscous liquid
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The test item was handled with all necessary protective clothing and all recommended safety and sterile measures were followed. The identity, composition stability and characteristics of the test item is the responsibility of the sponsor. No analysis was performed at GLR Laboratories Private Limited, to confirm it.

**Description of the test item**

The test item, Taglus PU FLEX Thermoforming Foils is a transparent disk with a diameter of 12.5 cm and thickness less than 0.5 mm. It is a surface device which comes in contact with mucosal membrane. The duration of contact is less than 24 hours (limited).

**TEST SYSTEM**

Species	<i>Oryctolagus cuniculus</i> (Rabbit)
Strain	New Zealand White
Weight range (g) (at the time of dosing)	2093.4 to 2368.5



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Sex	Male
Source	Animals were procured from the supplier approved by, the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) and were quarantined for 7 days. Supplier - VAB Biosciences #1-6-197/45/D, Bapuji Nagar, Musheerabad, Hyderabad-500020.
Number of animals used	6
Acclimatization period	6 days
Justification for animal use	Rabbits were selected because there is a large volume of background data on this species Recommended in ISO 10993-23:2021(E) standard as an appropriate species to evaluate skin irritation of medical devices and by various regulatory authorities.

The test system was approved by the Institutional Animal Ethics Committee (IAEC) of GLR Laboratories Private Limited.

**ANIMAL HUSBANDRY**

Test room no.	20
Test room temperature (°C)	18.1 to 21.9
Relative humidity (%)	37 to 58
Housing	Animals were housed individually in standard rabbit cages.
Method of identification	Animals were identified using cage cards indicating cage no., study no., species, strain, animal no., sex, body weight, dose, signature and individual numbers.
Feed	Commercial rabbit pellet feed Supplier - VRK Nutritional solutions D-47 & W-38, MIDC area, Miraj, Dist. Sangli- 416410, Maharashtra (India)
Water	Purified drinking water was provided <i>ad libitum</i>
Bedding material	No bedding material was used as rabbits are housed in stainless steel cages with mesh floors. A tray with sterilized paddy husk used to collect the excreta and urine was changed every day. This was not in contact with the rabbits directly.



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Supplier - M/S K. Dhandapani  
4/331, Old Mahabalipuram Road  
Kottivakkam, Chennai-600041  
Tamilnadu (India)

Photoperiod	12 h light and 12 h dark cycle
Contaminants	Contaminants, reasonably expected in feed or water supplies are not believed to influence the outcome of the study. Analysis of feed, water and bedding materials are carried out once in every 6 months and the results of the most recent analysis were placed in the study file.
Personnel	Appropriately qualified and trained personnel were involved in this study.
Selection of animals	Only healthy, previously unused and healthy young adults were selected for this study.

### TEST METHOD

#### Preparation of the test item and control item

The whole test item was extracted at a ratio of 6 cm<sup>2</sup>/mL (since the thickness of the test item is less than 0.5 mm) in polar solvent (physiological saline) as well as non-polar solvent (cottonseed oil) at 37 ± 1 °C for 72 h and 02 min under sterile conditions. Solvent controls were also subjected to same extraction conditions. Total surface area of the test item is 111 cm<sup>2</sup> (as calculated in our laboratory). This fulfils the requirements of ISO 10993-12:2012(E) and ISO 10993-12:2021(E).

Extract	Extraction vehicle	Surface area of the test item (cm <sup>2</sup> )	Volume of vehicle (mL)	Extract preparation start time	Extract preparation end time	Appearance of extracts
Extract 1	Physiological saline	111	18.5			Colourless clear liquid, no particulates*
Extract 2	Physiological saline	NA	10	09:00 a.m. on 03 Dec 2021	09:02 a.m. on 06 Dec 2021	Colourless clear liquid, no particulates
Extract 3	Cottonseed oil	111	18.5			Yellow viscous liquid, no particulates*
Extract 4	Cottonseed oil	NA	10			Yellow viscous liquid, no particulates

NA-Not applicable; Extraction time: 72 h and 02 min.

\*No change in colour of the extract, compared to extraction vehicle alone.

No changes were observed in the retrieved test item. No additional processing such as filtration, centrifugation, pH adjustments or any other processing were made. The extracts and solvent controls were transferred to sterile containers and stored at room temperature.

All extracts and solvent controls were used within 1 h and 23 min of preparation and were considered stable during this time.

### **Dosing procedure**

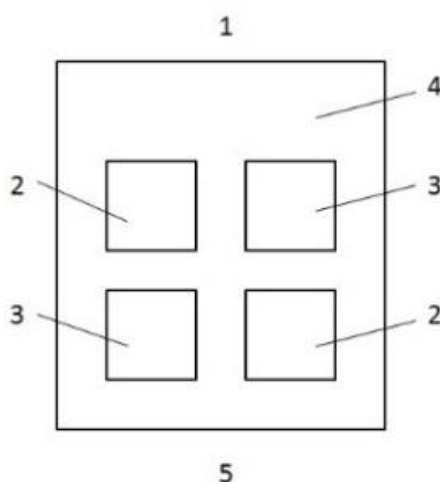
Justification for method of application Specified in ISO 10993-23:2021(E) standards, skin irritation in rabbit is recommended as a suitable method to determine the biocompatibility of medical devices.

The animals with healthy intact skin were selected for this study. About 16 h and 55 min prior to application, fur on all the rabbits were closely clipped off their backs for an area of 10 cm x 15 cm on both sides of the spinal cord.

### **Topical application**

Test item extract (0.5 mL) was loaded on the absorbent gauze patch measuring 6.25 cm<sup>2</sup> (2.5 cm x 2.5 cm) which was placed as such on to the fur clipped area of rabbit skin, in the dorsal region on the left cranial end and right caudal end. Similarly, 0.5 mL of solvent controls was loaded on the absorbent gauze (Make: The Ramaraju Surgical Cotton Mills; Batch. No: 578/19; Expiry Date: Jul 2022) measuring 6.25 cm<sup>2</sup> (2.5 cm x 2.5 cm) which was placed as such on to the fur clipped area of rabbit skin, the right cranial end and left caudal end as shown in the figure below.

The patches were loosely held in contact with the skin by semi-occlusive dressing with means of non-irritant adhesive tape (Make: 3M India Limited; Batch No.: R05190315; Expiry Date: Apr 2024) for the duration of 4 h. No test item extracts residues were observed. The test sites were marked with non- irritant permanent ink.



1. Cranial end; 2. Test site; 3. Control site; 4. Clipped dorsal region; 5. Caudal end.

Source: ISO 10993-23:2021(E)

## **OBSERVATIONS**

### **Mortality & Morbidity**

Animals were observed for mortality and morbidity daily throughout the experiment.

### **Body Weight**

Body weights of each animal were recorded prior to dosing and end of the experiment.

### **Clinical observations**

Animals were examined for signs of erythema and oedema. The responses were scored at 1 h, and then at 24 h, 48 h and 72 h following the patch removal.

### **Grading of skin reactions**

Animals were macroscopically examined for signs of erythema and oedema, visually with naked eyes. Skin reactions were graded and recorded at 1 h, and then at 24 h, 48 h and 72 h following the patch removal according to ISO 10993-23:2021(E).

Skin reactions were recorded at each examination as shown in the table below.

<b>Erythema and Eschar Formation</b>	
No erythema	0
Very slight erythema (barely perceptible)	1
Well defined erythema	2
Moderate erythema	3
Severe erythema (beet-redness) to eschar formation preventing grading of erythema	4
<b>Oedema Formation</b>	
No oedema	0
Very slight oedema (barely perceptible)	1
Well-defined oedema (edges of area well defined by definite raising)	2
Moderate oedema (raised approximately 1 mm)	3
Severe oedema (raised more than 1 mm and extending beyond exposure area)	4
<b>Maximum possible score for irritation: 8</b>	

Source: ISO 10993-23:2021(E)

In addition to the observation of irritation, all local toxic effects, such as defatting of the skin, and any systemic adverse effects (e.g., clinical signs of toxicity and body weight), were recorded.

## **DATA EVALUATION**

The skin irritation scores were evaluated in conjunction with the nature and severity of lesions, and their reversibility or lack of reversibility. The individual scores do not represent an absolute standard for the irritant properties of a material, as other effects of the test material are also evaluated.

After 72 h grading, all erythema grades plus oedema grades at 24 h, 48 h and 72 h were totalled separately for test item and control for each animal. The primary irritation score for an animal was calculated by dividing the sum of all the scores by 6 (two test/observation

sites, three time points). The primary irritation index (PII) of the test item and control was obtained by adding the scores of the individual animals and dividing it by the total number of animals. The results were evaluated by calculating the difference between the primary irritation score of control and test item.

Based on the observations and primary irritation response, the test item was categorised as per the primary irritation index (Appendix 2).

### **ACCEPTANCE CRITERIA**

The present assay is considered valid based on it meeting the following criteria:

1. Positive control trial conducted within the test facility indicated a clear positive result.
2. Solvent controls used in the study gave a mean irritation score of 0.

### **RESULTS**

#### **Mortality & Morbidity**

No mortality or morbidity occurred in any of the animals used in this study.

#### **Body Weight**

A gradual increase in body weight was observed in all the animals at the end of the experiment. Individual body weight of the animals is given in Table 1.

#### **Clinical observations**

No treatment related clinical signs of toxicity were observed in the experimental animals.

#### **Grading of skin reactions**

The individual score for erythema/eschar and oedema of the test site and control site after 1 h, 24 h, 48 h and 72 h following patch removal are given in Table 2 and Table 3 for all the animals. Mean irritation scores of grading and the difference in primary irritation index of test and control sites are given in Tables 4, 5 and Table 6.

#### **Euthanasia**

Animals were euthanized by overdose of thiopental sodium injection at the end of the experiment.

#### **Necropsy & Gross pathology**

None of the animals were found dead or in moribund condition and therefore, no necropsy and gross pathology were conducted.





## **CONCLUSION**

Based upon the results obtained in this study and in line ISO 10993-23:2021(E), the given test item, Taglus PU FLEX Thermoforming Foils, supplied by Vedia Solutions, is considered as a non-irritant under the conditions of the present study.

## **REFERENCES**

1. Biological Evaluation of Medical Devices - Part 1, Evaluation and Testing within a Risk Management Process, ISO 10993-1:2018(E).
2. Biological Evaluation of Medical Devices - Part 2, Animal Welfare Requirements, ISO 10993-2:2006(E).
3. Biological Evaluation of Medical Devices - Part 23, Tests for Irritation, ISO 10993-23:2021(E).
4. Biological Evaluation of Medical Devices - Part 12, Sample Preparation and Reference Materials, ISO 10993-12:2012(E).
5. Biological Evaluation of Medical Devices - Part 12, Sample Preparation and Reference Materials, ISO 10993-12:2021(E).
6. OECD Principles of Good Laboratory Practice. OECD Environmental Health and Safety Publications, Series on Principles of Good Laboratory Practice and Compliance Monitoring No. 1. ENV/MC/CHEM (98)17.
7. General Requirements for the Competence of Testing and Calibration Laboratories, ISO/IEC 17025:2017(E).
8. Use of International Standard ISO 10993-1, "Biological Evaluation of medical devices - Part 1: Evaluation and Testing within a risk management process". Guidance for Industry and Food and Drug Administration Staff. September 04, 2020.

**Table 1: Individual body weights**

Animal number	Sex	Individual body weights (grams)	
		At the time of dosing	At the end of experiment
1	Male	2180.3	2230.2
2		2222.6	2268.3
3		2368.5	2419.3
4		2093.4	2142.3
5		2332.6	2383.4
6		2295.0	2341.1

**Table 2: Individual grades of skin reactions**

	Observation Time (h)	Individual score																	
		Animal number 1						Animal number 2						Animal number 3					
		T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T	T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T	T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T
Erythema and Eschar formation	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	24	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	72	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Oedema formation	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	24	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	72	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

C<sub>1</sub>-First control site; C<sub>2</sub>-Second control site; T-Total (Sum of C<sub>1</sub> & C<sub>2</sub>)

T<sub>1</sub>- First test site; T<sub>2</sub>- Second test site; T-Total (Sum of T<sub>1</sub> & T<sub>2</sub>)

Source: ISO 10993-23:2021(E)

**Table 3: Individual grades of skin reactions**

	Observation Time (h)	Individual score																	
		Animal number 4						Animal number 5						Animal number 6					
		T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T	T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T	T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	T
Erythema and Eschar formation	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	24	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	72	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Oedema formation	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	24	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	48	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
	72	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

C<sub>1</sub>-First control site; C<sub>2</sub>-Second control site; T-Total (Sum of C<sub>1</sub> & C<sub>2</sub>)

T<sub>1</sub>- First test site; T<sub>2</sub>- Second test site; T-Total (Sum of T<sub>1</sub> & T<sub>2</sub>)

Source: ISO 10993-23:2021(E)

**Table 4: Calculation of primary irritation score at three time points**

	Observation Time (h)	Individual score								
		Animal number 1			Animal number 2			Animal number 3		
		Score	Total Score	PI Score	Score	Total Score	PI Score	Score	Total Score	PI Score
Test (T)	Erythema and Eschar formation	24	0		0			0		
		48	0		0			0		
		72	0	0	0	0	0	0	0	0
	Oedema formation	24	0		0	0	0	0	0	
		48	0		0			0		
		72	0		0			0		



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Control (C)	Erythema	24	0			0			0		
	and Eschar	48	0			0			0		
	formation	72	0	0	0	0	0	0	0	0	0
		24	0			0			0		
	Oedema	48	0			0			0		
	formation	72	0			0			0		

Total score = Sum of all the scores at test site (or) control site;

Primary Irritation (PI) Score = Total score divided by 6;

Source: ISO 10993-23:2021(E)

**Table 5: Calculation of primary irritation score at three time points**

Observation Time (h)			Individual score								
			Animal number 4			Animal number 5			Animal number 6		
			Score	Total Score	PI Score	Score	Total Score	PI Score	Score	Total Score	PI Score
Test (T)	Erythema	24	0			0			0		
	and Eschar	48	0			0			0		
	formation	72	0	0	0	0	0	0	0	0	0
		24	0			0			0		
	Oedema	48	0			0			0		
	formation	72	0			0			0		
Control (C)	Erythema	24	0			0			0		
	and Eschar	48	0			0			0		
	formation	72	0	0	0	0	0	0	0	0	0
		24	0			0			0		
	Oedema	48	0			0			0		
	formation	72	0			0			0		

Total score = Sum of all the scores at test site (or) control site;

Primary Irritation (PI) Score = Total score divided by 6;

Source: ISO 10993-23:2021(E)

**Table 6: Calculation for Primary Irritation Index and Primary Irritation difference by using Primary Irritation Score**

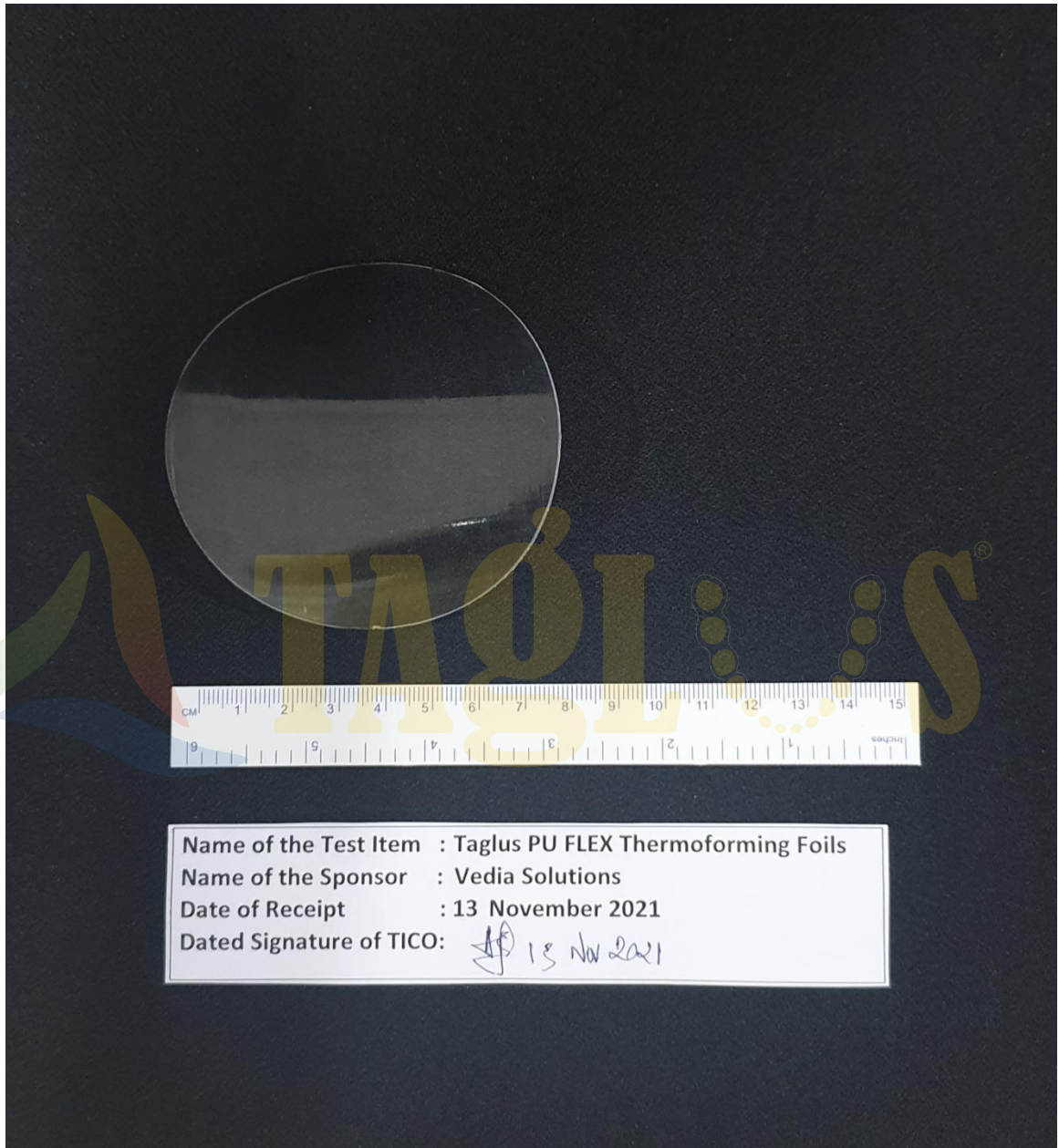
Animal number	1	2	3	PII*	PII difference <sup>#</sup>	4	5	6	PII*	PII difference <sup>#</sup>
Negative control site	0	0	0	0	0	0	0	0	0	0
Test item site	0	0	0	0		0	0	0	0	

\* Primary irritation index (sum of all primary irritation scores/number of animals)

<sup>#</sup> PII difference = PII of test site - PII of control site

Source: ISO 10993-23:2021(E)

**PHOTOGRAPH OF THE TEST ITEM**





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**APPENDIX 1**  
**CONCISE POSITIVE CONTROL STUDY DATA**

Study number	000/054
Study title	Skin Irritation Test in New Zealand White Rabbits
Study start date	27 September 2021
Experiment start date	28 September 2021
Experiment completion date	15 October 2021
Study completion date	20 October 2021

**INTRODUCTION**

Skin irritation is a key toxicity endpoint to assess biocompatibility of chemicals/medical devices. An assessment is made for testing the potential of the material under test to produce dermal irritation in rabbits following topical application.

**OBJECTIVE**

This skin irritation test was conducted to demonstrate the positive response of Sodium Lauryl Sulphate in New Zealand White Rabbits in compliance with ISO 10993-10:2010(E), ISO 10993-23:2021(E) and OECD 404:2015 guidelines.

**DETAILS OF TEST ITEM [Sodium Lauryl Sulphate]**

Appearance/Colour	Form: Powder, Colour: White
Manufacturer	Sigma Aldrich
CAS No.	151-21-3
Batch No.	0000009635
Manufacture Date	Not available
Expiry Date	August 2022
Concentration used in study	20% w/v

**METHODOLOGY**

This test was performed based on ISO 10993-10:2010(E), ISO 10993-23:2021(E) and OECD 404:2015 guidelines.

Two grams of Sodium Lauryl Sulphate was dissolved in distilled water and made up to 10 mL to obtain 20% w/v Sodium Lauryl Sulphate solution. Three male rabbits were clipped free of fur on dorsal side from an area of approximately 10 cm x 15 cm on both sides of the spinal cord approximately 16 h and 30 min prior to commencement of the experiment. The test item (0.5 mL) was applied onto the gauze measuring 6.25 cm<sup>2</sup> (2.5 cm x 2.5 cm) and placed on the test site in the dorsal region on the left cranial end and right caudal end of rabbit skin.

Similarly, 0.5 mL of the negative control (distilled water) was applied onto the gauze measuring 6.25 cm<sup>2</sup> (2.5 cm x 2.5 cm) and placed in the right cranial end and left caudal end on the control site.

The application sites were covered with a gauze patch (Make.: The Ramaraju Surgical Cotton Mills Limited; Batch No: 578/19; Expiry Date: July 2022) which was loosely held in contact with the skin by means of a suitable semi-occlusive dressing and non-irritant adhesive tape (Make.: 3M India Limited; Batch No.: R05190315; Expiry Date: April 2024) for all the animals. The patches were removed, 4 hours after the test item application and the test sites were marked with non-irritant permanent ink. No residues of the test item were found at the test site after patch removal.

## STUDY RESULTS

### Individual grades of skin reactions

	Observation Time (h)	Animal number 1							Individual score							Animal number 3						
		T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	C		T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	C		T <sub>1</sub>	T <sub>2</sub>	T	C <sub>1</sub>	C <sub>2</sub>	C	
Erythema and Eschar formation	1	1	1	2	0	0	0		0	1	1	0	0	0		0	1	1	0	0	0	
	24	1	2	3	0	0	0		2	2	4	0	0	0		2	2	4	0	0	0	
	48	1	2	3	0	0	0		2	2	4	0	0	0		1	2	3	0	0	0	
	72	2	2	4	0	0	0		2	2	4	0	0	0		2	2	4	0	0	0	
	Day 7	1	0	1	0	0	0		1	1	2	0	0	0		0	0	0	0	0	0	
Oedema formation	1	0	0	0	0	0	0		0	0	0	0	0	0		0	0	0	0	0	0	
	24	1	2	3	0	0	0		1	2	3	0	0	0		1	2	3	0	0	0	
	48	1	1	2	0	0	0		1	2	3	0	0	0		2	1	3	0	0	0	
	72	1	2	3	0	0	0		1	2	3	0	0	0		2	1	3	0	0	0	
	Day 7	0	0	0	0	0	0		1	0	1	0	0	0		0	0	0	0	0	0	

C<sub>1</sub>-First control site; C<sub>2</sub>-Second control site; C-Sum of C<sub>1</sub> & C<sub>2</sub>

T<sub>1</sub>- First test site; T<sub>2</sub>- Second test site; T-Sum of T<sub>1</sub> & T<sub>2</sub>

Source: ISO 10993-10:2010 (E) and ISO 10993-23:2021(E)

### Calculation of primary irritation score at three time points

Sites	Skin Reaction	Observation Time (h)	Individual score								
			Animal number 1			Animal number 2			Animal number 3		
			Score	Total Score	PI Score	Score	Total Score	PI Score	Score	Total Score	PI Score
Test (T)	Erythema and Eschar formation	24	3			4			4		
		48	3			4			3		
		72	4	18	3.0	4	21	3.5	4	20	3.3
	Oedema formation	24	3			3			3		
		48	2			3			3		
		72	3			3			3		
Control (C)	Erythema and Eschar formation	24	0			0			0		
		48	0			0			0		
		72	0	0	0	0	0	0	0	0	0
	Oedema formation	24	0			0			0		
		48	0			0			0		
		72	0			0			0		

Total score = Sum of all the scores at test site (or) negative control site;

Primary Irritation (PI) Score = Total score divided by 6;

Source: ISO 10993-10:2010 (E) and ISO 10993-23:2021(E)

**Calculation for Primary Irritation Index and Primary Irritation difference by using Primary Irritation Score**

Animal number	1	2	3	PII	PII difference
Negative control site	0	0	0	0	3.3
Test item site	3.0	3.5	3.3	3.3	

Primary irritation index (PII) = Sum of all primary irritation scores divided by 3

PII difference = PII of test site - PII of negative control site

Source: ISO 10993-10:2010 (E) and ISO 10993-23:2021(E)

## DISCUSSION

Based on the primary irritation index obtained, 20% w/v Sodium Lauryl Sulphate is considered as an irritant to rabbit skin. Given that the mucosal membranes are more prone to irritant effects of chemicals, than the skin, it can be considered that 20% Sodium Lauryl Sulphate may induce irritation in mucosal membranes. Therefore, no separate animal experiments were performed in view of 3R's principles of animal testing.

## CONCLUSION

Based on the results obtained, 20% w/v Sodium Lauryl Sulphate induced a primary irritation score of 3.3 and hence concluded as a moderate irritant under the conditions of the present study.

### Summary of Positive Control Trial (GLR Study number 000/054)

Study number	Study start date	Experiment start date	Experiment completion date	Study completion date	Agent used	Result
000/054	27 September 2021	28 September 2021	15 October 2021	20 October 2021	20% Sodium Lauryl Sulphate	Moderate irritant





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**APPENDIX 2**

**Primary Irritation Index (PII)**

Mean Score	Response category
0 to 0.4	Negligible
0.5 to 1.9	Slight
2 to 4.9	Moderate
5 to 8	Severe

Source: ISO 10993-23:2021(E)







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**RESPONSIBLE PERSONNEL**

Mr. G. Santhakumar, MSc, ERT	Study Director
Ms. S. Bharkavi, MSc	Study Scientist
Ms. T. Gayathri, MSc	Study Scientist
Dr. D. Yogaraj, MVSc	Study Scientist
Dr. S. Kavirajan, MVSc	Veterinarian
Dr. L. Mayavan, BVSc & AH	Animal House In-charge

**STATEMENT OF STUDY COMPLIANCE**

This study was performed in compliance with:

- OECD Principles of Good Laboratory Practice (revised 1997, issued January 1998) ENV/MC/CHEM (98) 17.
- US Food and Drug Administration's GLP regulations, 21 CFR 58 (subparts B to G and J).
- ISO/IEC 17025:2017(E) (general requirements for the competence of testing and calibration laboratories).

All procedures were performed in accordance with GLR Laboratories Private Limited Standard Operating Procedures (SOPs). The study was subjected to Quality Assurance evaluation by the GLR Laboratories Private Limited Quality Assurance Unit (QAU) in accordance with SOPs.

**STUDY PLAN AMENDMENT**

No study plan amendment was made during the conduct of the study.

**STUDY PLAN DEVIATION**

No study plan deviation occurred during the conduct of the study.

**ARCHIVE STATEMENT**

All primary data, or authenticated copies thereof a sample test item, study plan and the final report will be retained for a period of 9 years in the GLR Laboratories Private Limited archives after issue of the final report. At the end of the specified archive period the Sponsor will be contacted to determine whether the data should be returned, retained or



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destroyed on their behalf. Sponsors will be notified of the financial implications of each of these options at that time.

**DISTRIBUTION OF REPORTS**

Two originals of the study report are prepared and distributed as mentioned below:

1. Sponsor.
2. Archive (GLR Laboratories Private Limited).



**ANNEXURE 1**

  
**GOVERNMENT OF INDIA**  
**Department of Science and Technology**  
**National Good Laboratory Practice (GLP) Compliance Monitoring Authority (NGCMA)**

## Certificate of GLP Compliance

This is to certify that

**GLR Laboratories Private Limited**  
**444, Gokulam Street, Mathur**  
**Madhavaram, Chennai-600068 (Tamil Nadu), India**

is a GLP certified test facility in compliance with the NGCMA's Document No. GLP-101 "Terms & Conditions of NGCMA for obtaining and maintaining GLP certification by a test facility" and OECD Principles of GLP.

The test facility conducts the below-mentioned tests/ studies:

- **Toxicity Studies**
- **Mutagenicity Studies**

The specific areas of expertise, test items and test systems are listed in the annexure overleaf.

**Validity: March 13, 2020 – April 3, 2022**

Certificate No. : GLP/C-132A/2020  
Issue Date : 13-03-2020



  
**(Dr. Neeraj Sharma)**  
Head, NGCMA